

Nutritional and growth performance of feeding *Polyalthia longifolia* leaf meal as partial replacement of wheat offal in the diet of broiler chicks

Oluwafemi RA¹, Akinbisola SA², Alagbe JO³

^{1,2} Department of Animal Science, University of Abuja, Nigeria

³ Department of Animal Nutrition & Biochemistry, Sumitra Research Institute, Gujarat, India

Abstract

A total of two hundred and fifty (250) one day old (Ross 308) broiler chicks of mixed sex were used to determine the nutritional and growth performance of feeding *Polyalthia longifolia* leaf meal (PLM) as partial replacement of wheat offal in the diet of broiler chicks. Birds were divided to five treatments with five replicates of ten (10) birds in a completely randomized design. Treatment 1 (T1) was fed diet with no PLM, T2, T3, T4 and T5 were fed diet with PLM replacing wheat offal at 5 %, 10 %, 15 % and 20 % respectively. Results on proximate analysis of PLM revealed the presence of dry matter (91.40 %), crude protein (10.01 %), crude fibre (19.70 %), ether extract (0.18 %), ash (6.02 %) and energy (1510 kcal/kg). Mineral analysis showed that PLM contained calcium (71.22 mg/100g), phosphorus (67.12 mg/100g), potassium (11.23 mg/100g), iron (5.32 mg/100g), sodium (92.31 mg/100g) and magnesium (19.33 mg/100g). Qualitative phytochemical analysis revealed the presence of alkaloids (1.08 mg/100g), tannins (2.21 mg/100g), flavonoids (13.10 mg/100g), terpenoids (3.09 mg/100g) and phenols (2.28 mg/100g). Weight gain, Average daily feed intake and feed: gain were significantly influenced by PLM ($P < 0.05$). Birds in T1 had the highest mortality (3.00 %) while T2 recorded (1.26 %), none was recorded in T3, T4 and T5 ($P < 0.05$). It was concluded that PLM could be used to replace wheat offal up to 20 % without causing any negative effect on the general performance of birds.

Keywords: broiler chicks, *Polyalthia longifolia* leaf, phytochemicals, wheat offal

Introduction

Feed has been a major cost in modern broiler production, accounting about 70% of the total production cost (Onigemo, 2012) [38]. The increase in feed price may therefore imply in the increase in total production cost and thus decrease the profit margin of broiler industry. Attempt has recently been taken to reduce the cost of feed, including the incorporation of agro-industrial by-products in broiler diets as an energy source (Alu *et al.*, 2012) [35]. However, some limitations may exist when using the agro-industrial by-products as ingredients in broiler rations. The high and low contents of fibre and protein in the by-products may limit the digestibility and thus inclusion level of such by-products (Sugiharto *et al.*, 2018a). In addition to the agro-industrial by-products, the application of leaf meal as the ingredient in broiler feeds has also been conducted (Aletor, 2007) [33]. Compared to agro-industrial by-products, the content of crude protein in leaf meal is much higher (Etuk *et al.*, 2011) [34]. This may be beneficial in reducing the proportion of the conventional-expensive protein-rich feed ingredients in broiler rations.

It has been known that some particular foliage's contain a number of bioactive compounds that are beneficial for the health of chickens (Rama Rao *et al.*, 2019). These compounds include vitamins, phenolic acids, flavonoids, isothiocyanates, tannins as well as saponins (Olafadehan *et al.*, 2020) [27]. In this regard, the use of leaf meal in rations may not only reduce the cost of feeds, but also elicit the health-promoting effect on broiler chickens. Another way of making protein available to people in cheaper prices especially in developing countries is by the use of

agricultural by-products and tropical plants (leaf meal) which are not directly used by humans as food to feed livestock (Asar *et al.*, 2010) [30]. It has been reported that some leaf meal provides animals with necessary vitamins, minerals, oxycarotenoids and protein (Francis, 2010), for instance *Polyalthia longifolia* leaves are found to be loaded with minerals and vitamins. It also an excellent source of sulphur containing amino acids which are often limiting in most feedstuff used for feeding animals (Alagbe, 2007).

The aim of this experiment was to determine the nutritional and growth performance of feeding *Polyalthia longifolia* leaf meal (PLM) as partial replacement of wheat offal in the diet of broiler chicks.

Materials and Methods

Experimental site

The study was carried out at the poultry section of the Teaching and Research Farm of the Faculty of Agriculture University of Abuja, Abuja Nigeria. The study carried out lasted for 8 weeks. The study territory exist in the southern guinea savanna ecological zone of Nigeria, geographically situated within latitude 08^o25' and 9^o 20' N and longitude 06^o 45' and 07^o39' E.

Collection and Processing of *Polyalthia longifolia* Leaf

Fresh, healthy and mature *P. longifolia* leaves were obtained in Kuje, Kuje Area Council of the Federal Capital Territory. The leaves were washed with running tap water to remove the dirt's, it was later air dried separately until constant weights were obtained and made to meal using a hammer mill. The sample was later stored in an air tight container at 4°C for further analysis.

Animals and their management

Two hundred and fifty (250), Ross 308 one-day- old unsexed broiler chicks were randomly assigned to five dietary treatment groups of fifty birds per group with five replicates consisting of ten birds in a Completely Randomized Experimental Design. The birds were reared on a deep litter system and given anti-stress on arrival and antibiotics. Vaccination was administered according to the prevailing disease condition in the area. Feed and water were provided *ad libitum*. Feed intake, mortality were recorded daily while record of live weight were taken weekly throughout the experiment which lasted for 56 days.

Experimental diets and design

Birds were fed five experimental diets with *Polyalthia longifolia* leaf meal (PLM) replacing wheat offal at 0%, 5%, 10 %, 15 % and 20 % respectively in Completely Randomized Design.

Data collected

$$\text{Feed conversion ratio (FCR)} = \frac{\text{Dry Matter Intake (g)} \times 100}{\text{Live weight gain}}$$

$$\text{Average daily gain (ADG)} = \frac{\text{Final body weight} - \text{Initial body weight}}{\text{Total days of the experiment}}$$

Laboratory analysis

Proximate compositions of experiment diet and test materials were determined by using official method of analysis by AOAC (2000). Phytochemical composition of tannins, alkaloids, flavonoids, phenols and terpenoids were estimated using methods described by Harbone (1973), Odebiyi and Sofowora (1978) [23], Boham and Kocipai (1974) [21]. Minerals were determined using Atomic

Absorption Spectrophotometer (Model NF-123D, Punjab, India

Statistical analysis

All data were subjected to one -way analysis of variance (ANOVA) using SPSS (18.0) and significant means were separated using Duncan multiple range tests (Duncan, 1955) [16]. Significant was declared if $P \leq 0.05$.

Table 1: Composition of experimental diet (Broiler starter) 0 – 4 weeks

Ingredients	T ₁ 0 %	T ₂ 5 %	T ₃ 10 %	T ₄ 15%	T ₅ 20%
Maize	50.00	50.00	50.00	50.00	50.00
PLM	0.00	0.35	0.07	1.05	1.40
Wheat Offal	7.00	6.65	6.93	5.95	5.60
G/Cake 44%	5.50	5.50	5.50	5.50	5.50
Soya cake	30.05	30.05	30.05	30.05	30.05
Fish meal	2.00	2.00	2.00	2.00	2.00
Limestone	1.50	1.50	1.50	1.50	1.50
Bone meal	3.00	3.00	3.00	3.00	3.00
Salt	0.30	0.30	0.30	0.30	0.30
Vit TM Premix	0.25	0.25	0.25	0.25	0.25
Lysine	0.20	0.20	0.20	0.20	0.20
Methionine	0.20	0.20	0.20	0.20	0.20
Total	100.00	100.00	100.00	100.00	100.00
Calc. Analyses					
%CP	23.00	22.98	22.99	22.93	22.91
ME:Kcal/kg	2727	2728	2727	2730	2730
E:Prot ratio	118.59	118.73	118.62	119.02	119.16
EE %	4.59	4.58	4.59	4.56	4.55
CF %	4.71	4.74	4.72	4.80	4.83
Ca %	1.50	1.50	1.50	1.50	1.50
Avail P %	0.58	0.58	0.58	0.58	0.58
Ca: P ratio	2.60	2.60	2.60	2.60	2.60
Lysine %	1.43	1.44	1.43	1.44	1.44
Met + cys (%)	0.91	0.91	0.91	0.91	0.91

Table 2: Chemical composition of experimental diet (Broiler finisher) 5-8 weeks

Ingredients	T ₁ (0%)	T ₂ (5 %)	T ₃ (10 %)	T ₄ (15 %)	T ₅ (20 %)
Maize	60.00	60.00	60.00	60.00	60.00
PLM	0	0.25	0.26	0.75	1.01
Wheat Offal	5.05	4.80	4.90	4.30	4.04
G/Cake 44%	1.50	1.50	1.50	1.50	1.50
Soya bean meal	26.0	26.0	26.0	26.0	26.0
Fish meal (imported)	2.00	2.00	2.00	2.00	2.00
Limestone	1.50	1.50	1.50	1.50	1.50
Bone meal	3.00	3.00	3.00	3.00	3.00
Salt	0.30	0.30	0.30	0.30	0.30
Vit TM Premix	0.25	0.25	0.25	0.25	0.25
Lysine	0.20	0.20	0.20	0.20	0.20
Methionine	0.20	0.20	0.20	0.20	0.20
Total	100.00	100.00	100.110	100.00	100.00
Calc. Analyses					
%CP	20.07	20.06	20.08	20.03	20.01
ME:Kcal/kg	2853	2854	2855	2855	2856
E:Prot ratio	142.15	142.29	142.24	142.55	142.69
EE %	4.40	4.40	4.40	4.38	4.38
CF %	4.14	4.16	4.18	4.20	4.22
Ca %	1.48	1.48	1.48	1.48	1.49
Avail P %	0.57	0.57	0.57	0.57	0.57
Ca : P ratio	2.60	2.60	2.60	2.60	2.60
Lysine %	1.27	1.27	1.27	1.27	1.27
Met + cys (%)	0.83	0.83	0.84	0.84	0.84

*Premix supplied per kg diet :- Vit A, 7,000 I.U; Vit E, 5mg; Vit D3, 3000I.U, Vit K, 3mg; Vit B2, 5.5mg; Niacin, 25mg; Vit B12, 16mg; Choline chloride, 120mg; Mn, 5.2mg; Zn, 25mg; Cu, 2.6g; Folic acid, 2mg; Fe, 5g; Pantothenic acid, 10mg; Biotin, 30.5g; Antioxidant, 56mg

Table 3: Proximate composition of *Polyalthia longifolia* leaf meal

Parameters	Concentration
Dry matter (%)	91.40
Crude protein (%)	10.01
Crude fibre (%)	19.70
Ether extract (%)	0.18
Ash (%)	6.02
Energy (kcal/kg)	1510.0

Table 4: Mineral composition of *Polyalthia longifolia* leaf meal

Parameters	Composition (mg/100g)
Calcium	71.22
Phosphorus	67.12
Potassium	11.23
Iron	5.32
Sodium	92.31
Magnesium	19.33

Table 5: Phytochemical analysis of *Polyalthia longifolia* leaf meal

Parameters	Composition (mg/100g)	*Permissible range
Alkaloids	1.08	3.50
Tannins	2.21	7.50
Flavonoids	13.10	31.50
Terpenoids	3.09	-
Phenols	2.28	-

Table 6: Performance characteristics of broiler chicks fed different levels of PLM

Parameters	T1	T2	T3	T4	T5	SEM
IBW (g)	40.11	40.02	40.00	40.01	40.03	0.02
FWG (g)	1834.1 ^c	1994.6 ^b	2007.5 ^a	2100.2 ^a	2100.8 ^a	5.67
WG (g)	1794.0 ^c	1954.6 ^b	1967.5 ^b	2060.2 ^a	2061.0 ^a	8.11
TFI (g)	3800.8 ^a	3700.9 ^a	3687.5 ^b	3680.7 ^b	3680.0 ^b	12.5
ADF (g)	67.86 ^a	66.08 ^b	65.85 ^c	65.73 ^c	65.71 ^c	0.56
Feed:gain	2.11 ^a	1.89 ^b	1.87 ^b	1.79 ^c	1.78 ^c	0.02
Mortality (%)	3.00	1.26	-	-	-	0.01

Means in the same row with different superscripts differ significantly ($P < 0.05$)

Initial body weight (IBW); final body weight (FBW); weight gain (WG); average daily weight gain (ADWG); total feed intake (TFI)

Result and Discussion

Proximate composition of experimental diet

Table 1 and 2 shows the proximate composition of experimental diet. Broiler starter diet was fed between 0-4 weeks while finishers mash was fed between 5-8 weeks. It was observed that the crude fibre and ash content increased with increase in *Polyalthia longifolia* leaf meal inclusion. The values were highest in T4 and T5, crude protein and energy level fall within the ranges recommended by NRC (1994). The ether extract obtained from this study are in conformity with the values obtained by Aduku (2004) [19]; Olafadehan *et al.* (2020) [27] and Alagbe and Oluwafemi (2019) [24] the ash content obtained are in accordance with the reports of Alagbe *et al.* (2020).

Proximate and mineral composition of *Polyalthia longifolia* leaf meal (PLM)

Table 3 and 4 revealed the proximate and mineral composition of PLM. The proximate component contained dry matter (91.40 %), crude protein (10.01 %), crude fibre (19.70 %), ash (6.02 %), ether extract (0.18 %) and energy (1510.0 kcal/kg). This result conforms to the findings of

Ojewuyi *et al.* (2014) [13] but contrary to the findings of Alagbe (2017), these differences could be attributed to processing methods, geographical location and age of plants. Mineral composition of PLM revealed the presence of calcium (71.22 mg/100g), phosphorus (67.12 mg/100g), potassium (11.23 mg/100g), iron (5.32 mg/100g), sodium (92.31 mg/100g) and magnesium (19.33 mg/100g). In order of abundance sodium > calcium > phosphorus > magnesium > potassium > iron. According to Enin *et al.* (2014) sodium is responsible for the regulation of phosphorus, osmotic pressure, water balance, transmission of nerve impulses, transport of glucose and amino acid. Calcium plays a vital role in rigidity, strength and teeth formation (Alagbe, 2019; Ibrahim *et al.*, 2001). Potassium is vital in the regulation of water, electrolyte and functioning of the muscles (Musa *et al.*, 2020). Sodium is important in acid base balance (Uzama *et al.*, 2012) [7]. Magnesium is an activator of several key enzyme systems, including kinases, (ie. enzymes that catalyse the transfer of the terminal phosphate of ATP to sugar or other acceptors), mutases (transphosphorylation reactions), muscle ATPases, and the enzymes cholinesterase, alkaline phosphatase, enolase, isocitric dehydrogenase, arginase (magnesium is a component of the arginase molecule), deoxyribonuclease, and glutaminase (Alagbe and Motunrade, 2019). Iron is an essential for haemoglobin formation, normal functioning of the central nervous system, oxidation of carbohydrates, fats and protein (Uzama *et al.*, 2012) [7].

Table 5 revealed the phytochemical analysis of PLM. The sample contained alkaloids, tannins, flavonoids, terpenoids and phenols at 1.08 mg/100g, 2.21 mg/100g, 13.10 mg/100g, 3.09 mg/100g and 2.28 mg/100g respectively. According to Oluwafemi *et al.* (2020) [27], phytochemicals are bioactive chemicals or secondary metabolite which performs multiple biological roles. Terpenoids exhibit various pharmacological activities i.e., anti-inflammatory, anti-malarial, anticancer, antiviral and antibacterial roles (Mahato and Sen, 1997; Abdul *et al.*, 2013) [6]. Tannins are known to possess antimicrobial (Alagbe, 2017) and antiviral activity (Adisa *et al.*, 2010) [11]. Flavonoids and phenols have been reported to perform antioxidant activity; it has the ability to scavenge free radicals and prevent disease in the body of animals (Ojewuyi *et al.*, 2014) [13]. Alkaloids play a role of analgesics, antimicrobial and anti-inflammatory activities (Babajide *et al.*, 1999). However, all values reported for PLM were within lethal levels reported by Alagbe and Oluwafemi, 2019; Alagbe (2020) [24].

Performance characteristics of broiler chicks fed different levels of PLM

Table 6 revealed the performance characteristics of broiler chicks fed different levels of PLM. Initial body weight (IBW); final body weight (FBW); weight gain (WG); total feed intake (TFI) and average daily feed intake (ADF) ranged between 40.00 – 40.11 g, 1834.1 – 2100.2 g, 1794.0 – 2061.0 g, 3680.0 – 3800.8 g and 65.71 – 67.86 g were significantly different ($P < 0.05$) among the treatments. Birds in treatment T4 and T5 had the highest weight gain of 2060.2 g and 2061.0 g respectively, T1 had the lowest weight gain of 1794.0 g. The weight gain could be attributed to the presence of phytochemicals in PLM. Scientific reports showed that plant demonstrated a considerable antibacterial, antifungal, hepatoprotective, immunostimulatory, cytotoxic and antiviral activities (Olubukola *et al.*, 2020; Abdul *et al.*,

2013, Alagbe, 2017, 2019; Onasanwo *et al.*, 2011; Wakimoto *et al.*, 2008) [6, 3]. The results obtained is in conformity with the findings of Mohammed *et al.* (2016); Obajuluwa *et al.* (2020) when Yohimbe (*Pausynistalia yorimbe*) and larvacide was feed to broiler chickens. T3, T4 and T5 recorded no mortality probably due to the presence of flavonoids in PLM; similar result was recorded by Alagbe (2017) when PLM was fed to broiler chicks as a phytogetic feed additive. T4 and T5 had the best feed conversion ratio of 1.79 and 1.78; this result is in agreement with the reports of Adjei *et al.* (2015) when allacin was fed to broiler chicks.

Conclusion

There are larger numbers of leaf meals with enormous potentials in Nigeria and one of such is *P. longifolia* leaf. *P. longifolia* leaf has high energy, vitamins and antimicrobial activities which makes it a good replacement for other feed stuffs. The experiment has clearly shows that PLM can be partially used to replace wheat offal up to 20 % without any negative effect on the growth and health of birds.

References

- Adjei MB, Atuahene CC, Attah-Kotoku V. The response of broiler chickens to dietary inclusion of allacin. *Journal of Anim. Sci. Adv.* 2015; 5(5):1295-1301.
- Mohammed IE, Mosad AS, Mohammed A, Adel H. *Alexandria Journal of Veterinary Sciences.* 2016; 49(2):50-64.
- Onasanwo SA, Saba AB, Oridupa OA, Owoyele BV. Anti-nociceptive and antiinflammatory properties of ethanolic extract of *Lagnenaria breviflora* fruit in rat and mice. *Niger J Physiol. Sci.* 2011; 26:71-76.
- Wakimoto N, Yin D, Kelly J, Said J. Cucurbitacin B has a potent antiproliferative effect on breast cancer cells in vitro and in vivo. *Cancer Sci.* 2008; 99:1793-1797.
- Olubukola SO, Anthony JA, Adewale A. Sub-chronic administration of methanolic whole fruit extract of *Lagnenaria breviflora* induces mild toxicity in rats. *Pharmacognosy Mag.* 2020; 11:516-521.
- Abdul W, Mehreen G, Syed BJ, Mohammad N, Ajmal K. Phytochemical analysis of medicinal plants occurring in local area of Mardan. *Biochemistry and Analytical Biochemistry.* 2013; 2(114):1-4.
- Uzama D, Bwai MD, Sunday AT. The phytochemicals, proximate and elemental analysis of *Securinega virosa* leaf extract. *Research Journal in Engineering and Applied Sciences.* 2012; 1(6):351-354.
- Enin GN, Antia BS, Enin FG. Chemical assessment of the proximate, mineral and antinutrients composition of *Sida acuta* leaves. *Organic Chemistry.* 2014; 7(14):24654-24660.
- Babajide SO, Oluwalana SA, Ajala MO, Folarin MO. Phytochemical screening of seeds of *Acacia nilotica*. *The Boprospector.* 1999; 1(2):27-31.
- Ibrahim NDG, Abdulrahman EM, Ibrahim G. Elemental analysis of the leaves of *Vernonia amydalina* and its biological evaluation in rats. *Nig. J. Natural Products and Medicine.* 2001; 5:13-17.
- Adisa RM, Choudhary EA, Adenoye GA, Olorunsogo OO. Hypoglycaemic and biochemical properties of *Cnestis ferruginea*. *African journal of Complementary and Alternative Medicine.* 2010; 7:185-194.
- AOAC. Association of Official Analytical Chemists. *Official Methods of Analysis 19th Edition* Washington, D.C, 2000, Pages 69-77.
- Ojewuyi OB, Ajiboye TO, Adebajo EO, Balogun A, Mohammed AO. Proximate composition, phytochemical and mineral contents of young and mature *Polyalthia longifolia* Sonn.leaves Fountain *Journal of Natural and Applied Sciences.* 2014; 3(1):10-19.
- Obajuluwa OV, Sanwo KA, Egbeyale LT, Fafiolu OA. Performance, blood profile and gut morphometry of broiler chickens fed diets supplemented with Yohimbe (*Pausynistalia yohimbe*) and Larvacide. *Journal of Veterianary and Animal Science,* 2020.
- Alagbe JO. Effect of dietary inclusion of *Polyalthia longifolia* leaf meal as phytobiotic compared with antibiotics on the nutrient retention, immune response and serum biochemistry of broiler chicken. *Greener Journal of Agricultural Sciences.* 2017; 7(3):74-81.
- Duncan DB. Multiple range and multiple F-test. *Biometrics.* 1955; 11(1):1-42.
- National Research Council. Nutrient requirement of poultry 9th Rev Edn, Washington D.C. National Academy Press, 1994.
- Alagbe JO. Effect of dietary inclusion of *Polyalthia longifolia* leaf meal as phytobiotic compared with antibiotics on performance, carcass characteristics and haematology of broiler chicken. *Scholarly Journal of Agricultural Science.* 2017; 7(3):68-74.
- Aduku AO. Animal nutrition in the tropics: Feeds and feeding in monogastric and ruminant nutrition. *Journal of Applied Poultry Research.* 2004; 13:628-638.
- Alagbe JO. Haematology, serum biochemistry, relative organ weight and bacteria count of broiler chicken given different levels of *Luffa aegyptiaca* leaf extracts. *International Journal of Advanced Biological and Biomedical Research.* 2019; 7(4):382-392.
- Boham BA, Kocipai AC. Flavonoids and condensed tannins from leaves of Hawaiian *vaccinium vaticulatum* and *V. calycinium*. *Pacific Sci.* 1974; 48:458-463.
- Harborne JD. *Phytochemical methods: A guide to modern techniques of plant analysis.* Chapman and Hall, London, 1973, 279.
- Odebiyi A, Sofowora AE. Phytochemical Screening of Nigerian Medicinal Plant. Part III, *Lloydia.* 1978; 41:234-246.
- Oluwafemi RA, Isiaka Olawale, Alagbe JO. Recent trends in the utilization of medicinal plants as growth promoters in poultry nutrition- A review. *Research in: Agricultural and Veterinary Sciences.* 2020; 4(1):5-11.
- Alagbe JO, Sharma D, Xing Liu. Effect of aqueous *Piliostigma thonningii* leaf extracts on the haematological and serum biochemical indices of broiler chicken. *Noble International Journal of Agriculture and Food Technology.* 2019; 1(2):62-69.
- Alagbe JO, Sharma R, Eunice Abidemi Ojo, Shittu, MD, Bello Kamoru A. Chemical evaluation of the proximate, mineral, vitamins and phytochemical analysis of *Daniellia oliveri* stem bark. *International Journal of Biological and Chemical Studies.* 2020; 2(1):16-22
- Olafadehan OA, Oluwafemi RA, Alagbe JO. Performance, haemato-biochemical parameters of

- broiler chicks administered Rolfe (*Daniellia oliveri*) leaf extract as an antibiotic alternative. *Advances in Research and Reviews*, 2020, 1:4.
28. Alagbe JO. Performance, hematology and serum biochemical parameters of weaner rabbits fed different levels of fermented *Lagenaria brevifera* whole fruit extract. *Advances in Research and Reviews*, 2020, 1:5.
29. Alagbe JO. Performance, blood profile and carcass evaluation of growing grass cutters fed diets supplemented with matured *Polyalthia longifolia* leaf meal. *Scholarly Journal of Agricultural Science*. 2017; 7(2):44-49
30. Asar OP, Ghosh Bidnus, Kanti Das, Soroj Chatterjee, Goutam Chandra. Antibacterial potentiality and phytochemical analysis of mature leaves of *P. longifolia*. *The South Pacific Journal of Natural Science*, Vol 26, 2008.
31. Alagbe JO, Agubosi OCP, Ajagbe AD, Shittu MD, Akintayo Balogun OM. Performance, haematology and serum biochemical parameters of growing grass cutters fed *Phyllanthus amarus* and *Piliostigma thonningii* leaf meal mixture as partial replacement for Soya bean meal. *United International Journal for Research and Technology*. 2020; 2(1):14-23.
32. Maroyi Abbasi, Alireza Seidavi, Wuyi Liu, Leila Asadpour. Investigation on the effect of different levels of dried sweet orange pulp on the performance, carcass characteristics, physiological and biochemical parameters of chickens. *Saudi Journal of Biological Sciences*. 2015; 22:139-146.
33. Aletor VA. Plant and animal protein alternatives for poultry production in Nigeria. Paper presented at the Department of Animal Science, University of Ibadan. Poultry Association of Nigeria, Oyo State collaborative workshop on alternative feedstuffs for the Nigeria poultry industry, 2007.
34. Etuk EB, Udegbonam AC, Emenalom OO, Esonu BO. Effect of partial replacement of maize with 2:1:1 combination of plantain peels, yam peels and palm kernel cake in broiler starter diet. *Proceedings of the 36th Annual Nigerian Society for Animal Production Conference*, Abuja, Nigeria, 2011, 408-410.
35. Alu SE, Kaankuka FG, Kapechi MS. Effects of replacing maize with sugarcane scrapings meal (SCSM) on the haematological parameters and serum biochemical variables of broiler finisher birds. *Proceedings of the 37th Annual Nigerian Society for Animal Production Conference*, Makurdi, Nigeria, 2012, 334-337.
36. Olafadehan OA, Oluwafemi RA, Alagbe JO. Carcass quality, nutrient retention and caeca microbial population of broiler chicks administered Rolfe (*Daniellia oliveri*) leaf extract as an antibiotic alternative. *Journal of Drug Discovery*. 2020; 14(33):146-154.
37. Alagbe JO, Oluwafemi RA. Performance and haematological parameters of broiler chicks gives different levels of dried lemon grass (*Cymbopogon citratus*) and garlic (*Allium sativum*) extract. *Research in: Agricultural and Veterinary Sciences*. 2019; 3(2):102-111.
38. Onigemo MA, Agbalaya KK, Tijani LA, Asafa AR, Anjola OAJ, Agbaye FP, *et al.* Performance of growing pullets fed with diets containing gala waste ® as a substitute for maize. *Proceedings of the 37th Annual Nigerian Society for Animal Production Conference*, Makurdi, Nigeria, 2012, pp. 333-336.
39. Musa Bashir, Alagbe JO, Adegbite Motunrade Betty, Omokore EA. Growth performance, caeca microbial population and immune response of broiler chicks fed aqueous extract of *Balanites aegyptiaca* and *Alchornea cordifolia* stem bark mixture. *United Journal for Research and Technology*. 2020; 2(2):13-21.